

Genome characteristics of cluster EA *Microbacterium* bacteriophages HungryHenry, CaptainRex, and ChikPic isolated from soil in Tennessee

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Abstract

Bacteriophages HungryHenry, CaptainRex, and ChikPic were isolated from soil collected in Tennessee using the bacterium *Microbacterium foliorum*. These bacteriophages have genomes that are 41,516 bp, 39,941 bp, and 40,333 bp in length and encode 62, 61, and 63 putative genes, respectively. Based on gene content similarity, all three bacteriophages are assigned to actinobacteriophage cluster EA (subclusters EA1, EA2 and EA5, respectively).

Bacteriophage	HungryHenry	CaptainRex	ChikPic
Collection site GPS coordinates	36.532708 N, 87.351642 W	36.5311 N, 87.357381 W	36.58067 N, 87.45902 W
Collection site ambient temperatures	23° C	22° C	20° C
Number of 150-base sequencing reads	242,235	1,584,131	153,296
Sequencing coverage (fold)	875	152	415
Genome size (bp)	41,516	39,941	40,333
GC content (%)	63.4	63.5	62
Genome termini	Circularly permuted	Circularly permuted	Circularly permuted
Number of predicted genes	62	61	63
Number of genes with assigned functions	25	26	25
Number of tRNA genes	0	1	0
Subcluster	EA1	EA5	EA2

Table 1. Summary of relevant isolation parameters and genome characteristics of EA cluster bacteriophages HungryHenry, CaptainRex, and ChikPic:

All bacteriophages were found around Clarksville, Tennessee

Description

To expand our understanding of actinobacteriophage diversity and evolution, we described the isolation and characterization of three bacteriophages using *Microbacterium foliorum* (*M. foliorum*) NRRL B-24224 (Jacobs-Sera et al., 2020; Hatfull, 2020; Markov et al., 2022; Russell et al., 2019).

Bacteriophages HungryHenry, CaptainRex, and ChikPic were isolated from soil collected in Tennessee using standard methods (Zorawik et al., 2024). Collection site GPS coordinates and ambient temperatures are provided in Table 1. Briefly, soil samples were incubated for 2 hours in PYCa (peptone yeast calcium) liquid medium. The suspensions were then filtered (0.22- μ m pores), and the resulting filtrates were inoculated with *M. foliorum* and incubated with shaking (250 rpm) at 30°C for 2 days. An aliquot for each culture was then filtered and plated with PYCa top agar overlays containing *M. foliorum*. Following incubation at 30°C for 48 hours, representative plaques were selected for each sample and bacteriophages were purified through two additional rounds of plating. Bacteriophages for all three samples formed turbid plaques.

DNA was isolated from bacteriophage lysates using the Wizard DNA Clean-Up Kit (Promega, Madison, WI) and enzymatically sheared for sequencing using the Ultra II Library Kit (NEB, Ipswich, MA). An Illumina MiSeq instrument (v3 reagents) was used for DNA sequencing to yield 150-base single-end reads that were assembled using Newbler v2.9 (Russell, 2018) and checked for genomic termini and completeness using Consed v29, as previously described (Russell, 2018). Sequencing results and genome characteristics are provided in Table 1.

Bacteriophage genome annotation was conducted using DNA Master v5.23 (Pope et al., 2018), Glimmer v3.02 (Delcher et al., 1999), GeneMark v2.5p (Besemer and Borodovsky 2005), Aragorn v1.2.41 (Laslett et al., 2004), Phamerator v393.0 (Cresawn et al., 2011), PECAAN v20211202.0 (<https://blog.kbrinsgd.org/>), BLASTp (Altschul et al., 1990) against the NCBI non-redundant and Actinobacteriophage databases, tRNAscan-SE v2.0 (Lowe, 2016), HHPred v3.2 (Söding et al., 2005) against the PDB_mmCIF70, Pfam v.37.0, and NCBI Conserved Domains databases v3.19. Genes start locations were established using Starterator v485.0 (<http://phages.wustl.edu/starterator>). Except where otherwise noted, default parameters were used. The number of genes predicted along with putative functional assignments are presented in Table 1. Using the gene content similarity of at least 35% to bacteriophages in the Actinobacteriophage database, HungryHenry, CaptainRex, and ChikPic were all assigned to actinobacteriophage cluster EA, subclusters EA1, EA5, and EA2, respectively (Pope, 2017; Gauthier and Hatfull 2023). When compared to other EA bacteriophages described by Jacobs-Sera et al. (Jacobs-Sera et al., 2020) and in phagesdb.org (<https://phagesdb.org/>), CaptainRex (jointly with bacteriophage Librie) was found to have the shortest genome of EA5 subcluster. CaptainRex also had the lowest GC content of EA5 bacteriophages, and ChikPic was found to be one of two EA2 bacteriophages without an identifiable tRNA. Transmission electron micrographs for previously characterized cluster EA bacteriophages suggests that all three the bacteriophages are likely to have siphoviral morphologies (Jacobs-Sera et al., 2020).

Data availability

GenBank accession numbers for HungryHenry, CaptainRex, and ChikPic are [PP978832](#), OQ938588, and OR553911 respectively. SRA accession numbers are [SRX26311144](#), [SRX14443488](#), and [SRX26311141](#) respectively.

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References

- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *Journal of Molecular Biology* 215: 403-410. DOI: [10.1016/S0022-2836\(05\)80360-2](#)
- Besemer J, Borodovsky M. 2005. GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. *Nucleic Acids Research* 33: W451-W454. DOI: [10.1093/nar/gki487](#)
- Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. *BMC Bioinformatics* 12: 10.1186/1471-2105-12-395. DOI: [10.1186/1471-2105-12-395](#)
- Delcher A. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Research* 27: 4636-4641. DOI: [10.1093/nar/27.23.4636](#)
- Gauthier CH, Hatfull GF. 2023. PhamClust: a phage genome clustering tool using proteomic equivalence. *mSystems* 8: 10.1128/msystems.00443-23. DOI: [10.1128/msystems.00443-23](#)
- Gordon D, Abajian C, Green P. 1998. *Consed*: A Graphical Tool for Sequence Finishing. *Genome Research* 8: 195-202. DOI: [10.1101/gr.8.3.195](#)
- Hatfull GF. 2020. Actinobacteriophages: Genomics, Dynamics, and Applications. *Annual Review of Virology* 7: 37-61. DOI: [10.1146/annurev-virology-122019-070009](#)
- Jacobs-Sera D, Abad LA, Alvey RM, Anders KR, Aull HG, Bhalla SS, et al., Hatfull. 2020. Genomic diversity of bacteriophages infecting *Microbacterium* spp. *PLOS ONE* 15: e0234636. DOI: [10.1371/journal.pone.0234636](#)
- Laslett D. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Research* 32: 11-16. DOI: [10.1093/nar/gkh152](#)
- Lowe TM, Chan PP. 2016. tRNAscan-SE On-line: integrating search and context for analysis of transfer RNA genes. *Nucleic Acids Research* 44: W54-W57. DOI: [10.1093/nar/gkw413](#)
- Markov SA, Arms NL, Boyce KJ, Cardona Pendleton MR, Couch AM, Duncan LE, et al., Zimmer. 2022. Complete Genome Sequence of *Microbacterium foliorum* Bacteriophage Librie. *Microbiology Resource Announcements* 11: 10.1128/mra.00918-22. DOI: [10.1128/mra.00918-22](#)
- Pope WH, Mavrich TN, Garlena RA, Guerrero-Bustamante CA, Jacobs-Sera D, Montgomery MT, et al., Hatfull. 2017. Bacteriophages of *Gordonia* spp. Display a Spectrum of Diversity and Genetic Relationships. *mBio* 8: 10.1128/mbio.01069-17. DOI: [10.1128/mBio.01069-17](#)
- Pope WH, Jacobs-Sera D. 2017. Annotation of Bacteriophage Genome Sequences Using DNA Master: An Overview. *Methods in Molecular Biology, Bacteriophages* : 217-229. DOI: [10.1007/978-1-4939-7343-9_16](#)
- Russell DA. 2017. Sequencing, Assembling, and Finishing Complete Bacteriophage Genomes. *Methods in Molecular Biology, Bacteriophages* : 109-125. DOI: [10.1007/978-1-4939-7343-9_9](#)
- Russell DA, Garlena RA, Hatfull GF. 2019. Complete Genome Sequence of *Microbacterium foliorum* NRRL B-24224, a Host for Bacteriophage Discovery. *Microbiology Resource Announcements* 8: 10.1128/mra.01467-18. DOI: [10.1128/MRA.01467-18](#)
- Soding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. *Nucleic Acids Research* 33: W244-W248. DOI: [10.1093/nar/gki408](#)
- Zorawik M, Jacobs-Sera D, Freise AC, SEA-PHAGES, Reddi K. 2024. Isolation of Bacteriophages on Actinobacteria Hosts. *Methods in Molecular Biology, Phage Engineering and Analysis* : 273-298. DOI: [10.1007/978-1-0716-3798-2_17](#)

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